Basic Science for Clinicians

Vascular Smooth Muscle Cell Proliferation in Restenosis

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Current therapeutic approaches to restore blood flow in stenotic blood vessels involve the use of percutaneous devices and coronary bypass surgery. In all procedures that disrupt the normal integrity of the blood vessels, there is an increased incidence of vessel luminal narrowing, termed restenosis. Restenosis, arbitrarily defined as greater than 50% narrowing of vessel diameter compared with the reference vessel, is modulated by genetic background and diseases that affect the cardiovascular system, including diabetes, hypertension, and hypercholesterolemia. In the 1970s, Andreas Gruntzig pioneered the use of transluminal dilatation of coronary arteries for symptomatic coronary artery disease and reported a 19% rate of restenosis (6 of 32 patients).1-3 Subsequent studies demonstrated a restenosis rate of approximately 33%.4 More than 25 years later, despite pharmacological and mechanical approaches to reduce the incidence of restenosis, it remains a significant problem, especially in high-risk patient groups, limiting overall success.

Restenosis after percutaneous intervention is characterized by platelet aggregation, release of growth factors, inflammatory cell infiltration, medial smooth muscle cell proliferation and migration, and extracellular matrix remodeling. The vascular response to injury depends not only on the cells within the vessels but is also modulated by circulating bone marrow-derived cells. Understanding the molecular mechanisms underlying the physiological healing response and the pathological restenosis response has been the focus of extensive investigations, which have led to the development of novel approaches to control the pathological formation of the neointima. In this review, we will focus on some of the molecular mechanisms responsible for the abnormal neointimal hyperplasia, specifically focusing on cell cycle and microRNA (miRNA) in the vascular smooth muscle.

Pathophysiology of Restenosis

Acute occlusion at the percutaneous intervention site within hours to days after the procedure is usually caused by an intimal flap, thrombus formation, subintimal hemorrhage extending into the media, or elastic recoil of the overstretched vessel wall. The incidence of acute occlusion after percutaneous intervention has been minimized by the use of intravascular stents and aggressive antiplatelet therapy during the periprocedural period. Nevertheless, the overall rate of restenosis with bare-metal stents is approximately 20%.5 Drug-eluting stents have reduced the rate of restenosis, in most studies to <10%.

Animal models and human postmortem studies have shown that the mechanism of intimal hyperplasia is similar to wound healing.6 The wound-healing process can be divided into 3 phases: inflammatory phase (hours to days), granulation or cellular proliferation phase (days to weeks), and an extracellular matrix remodeling phase (months).6,7 After injury, there is deposition of platelets that release platelet-derived growth factor (PDGF) and other mitogenic factors that penetrate the vascular wall and trigger medial vascular smooth muscle cell proliferation and migration. The platelets also release chemokines, which initiate an inflammatory response. Over the first 2 weeks after injury, the vascular smooth muscle cells multiply 3 to 5 times, accounting for 90% of the ultimate intimal proliferation.

Endothelial Cells in Restenosis

A normal functioning endothelium is very important because it promotes vasodilatation and suppresses intimal hyperplasia by inhibiting thrombus formation, inflammation, and smooth muscle proliferation and migration. The endothelium provides a selectively permeable barrier that protects against circulating growth factors.8 Endothelial denudation and medial wall injury are the initial effects of balloon- and/or stent-induced injury and are important triggers of the wound “healing” program.

Vascular Smooth Muscle Cells

Vascular smooth muscle cells retain remarkable plasticity during postnatal development and can undergo dedifferentiation to a synthetic phenotype.9 This probably represents a survival advantage because it enables the efficient repair of the vasculature after injury. As in many evolutionarily conserved processes, these properties can be disadvantageous and can predispose to abnormal responses after injury, contributing to restenosis. Vascular smooth muscle progenitor cells have been identified in the bone marrow (multipotent vascular stem cell progenitors and mesenchymal stem cells), in the circulation (circulating progenitor cells), in the vessel wall (resident progenitors cells and mesangioblasts), and in extravascular sites.10

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Mechanisms of Smooth Muscle Cell Proliferation: The Cell Cycle

Smooth muscle cells progress through DNA replication and mitosis in a regulated series of cell-cycle events.17–20 Under normal conditions, quiescent smooth muscle cells are maintained in a nonproliferative phase (G0) (Figure 1). After vessel injury, smooth muscle cells enter a gap phase (G1), at which time factors necessary for DNA replication in the subsequent synthetic phase (S) are produced. After S phase, the cells enter another gap phase (G2), when proteins are synthesized for mitosis (M phase). Restriction points (R) at the G1-to-S and G2-to-M junctions ensure orderly progression through the cell cycle.

Growth factors, such as PDGF, basic fibroblast growth factor, and insulin-like growth factor-1, stimulate cells to enter the cell cycle and propel them to reach the R point in the late G1 phase. Beyond this, cell cycle progression from the G2 phase to the M phase does not require further growth factor stimulation. On binding to their respective cell surface tyrosine kinase receptors, growth factors trigger cell cycle entry by transactivating nuclear factors such as c-fos and c-myc. These nuclear factors act as transcriptional factors and increase the expression of various cell cycle regulatory proteins.

The cell cycle is coordinated by the expression and activities of regulatory proteins. Cyclins and their respective cyclin-dependent kinases (CDK) form distinct complexes and are positive regulators of cell cycle progression. CDK phosphorylation of the retinoblastoma gene product (Rb) at the R point at G1-S junction is an important step in the progression through the cell cycle.18 Rb ordinarily binds to and inactivates the transcription factor E2F to maintain the cell in a quiescent state. Phosphorylation of Rb in late G1 releases E2F, which in turn enhances the expression of genes encoding regulatory proteins necessary for cell cycle progression through S, G2, and M.21

The CDK inhibitors (CDKIs) are critical negative regulators of the cell cycle.22 The CDKIs are structurally divided into 2 different families: the INK4 family (p14, p15, p16, p18, and p19) and the KIP/CIP family (p21, p27, and p57). For example, p27Kip1, which binds to and inhibits phase G1 cyclin/CDK complexes, is maintained at high levels in quiescent cells and declines to permit cell cycle progression. In the later phases of arterial healing, p27Kip1 is upregulated, which is associated with inhibition of cellular proliferation.23 Another CDKI, p21Cip1, is present at low levels in G0, accumulates in late G1, and provides counterbalance to increased cyclin/CDK activities. Apart from CDKIs, other indirect cell cycle inhibitors also contribute to the regulation of mitosis.

Overexpression of p27Kip1 results in cell cycle arrest in the G1 phase.24 Gene transfer of p27Kip1 or p21Cip1 into balloon-injured vessels significantly reduced vascular smooth muscle cell proliferation and neointimal formation.23,25 Conversely, a reduction in p27Kip1 levels (caused by gene deletion in mice) resulted in increased neointimal formation and inflammatory cell accumulation after mechanical vascular injury.26

Mechanisms of Smooth Muscle Cell Migration

In the uninjured vessel, vascular smooth muscle cells are nonmigratory because of a combination of several factors including the relative absence of stimulatory factors, their quiescence from a proliferative standpoint, and because the matrix is highly adhesive.27 There are many promigratory and antimigratory molecules, including small biogenic amines, peptides growth factors, cytokines, and extracellular matrix components.27 Blood flow, shear stress, and matrix stiffness

Figure 1. Schematic of cell cycle. Cell cycle progression is dependent on the regulated expression and activation of a set of kinases (cyclin-dependent kinases, cdk) and cyclins, which phosphorylate proteins that regulate cell growth including the retinoblastoma protein (pRb indicates unphosphorylated; ppRb, phosphorylated form) at the restriction point (R) at the G1/S junction. Cdk inhibitors p27Kip1 and p21Cip1 delay cell cycle progression by inhibiting cdk activity.
can also affect the migration of vascular smooth muscle. Vascular smooth muscle cell migration begins with the stimulation of cell surface receptors that activate signal transduction pathways, which trigger remodeling of the cytoskeleton, changes in the adhesiveness of the smooth muscle cell to the matrix, and activation of the motor proteins. Vascular smooth muscle cells extend lamellipodia toward the stimulus through actin polymerization. Focal contacts form just behind the leading edge to increase adhesion of the cell membrane to the matrix. Vascular smooth muscle cells also must degrade the focal contacts in the trailing edge. The actin cytoskeleton is regulated by numerous signaling pathways and molecules, including trimeric G proteins, small G proteins, lipid kinases, Ca\(^{2+}\)-dependent kinases, Rho kinase, and mitogen-activated protein kinases. Promigratory stimuli that increase myosin II.30 Vascular smooth muscle cell migration is also linked to the cell cycle and proliferation. Smooth muscle cells can migrate when in the G1 phase but not in later phases of the cell cycle.31 p21\(^{Cip1}\) and p27\(^{Kip1}\) are not only negative regulators of the cell cycles but also have important inhibitory effects on vascular smooth muscle migration. Overexpression of p27\(^{Kip1}\) and p21\(^{Cip1}\) reduced human vein endothelial cell and vascular smooth muscle migration in vitro.32,33 p27\(^{Kip1}\) inhibits the cellular changes such as lamellipodia formation and reorganization of actin filaments and focal adhesions that are associated with vascular smooth muscle cell migration.34 A p27\(^{Kip1}\) mutant that lacks CDK inhibitory activity failed to inhibit the proliferation and migration of vascular smooth muscle cells.34 p27\(^{Kip1}\) also affects the sensitivity to rapamycin’s antimigratory effects in both vascular smooth muscle and endothelial cells (see below).

**Role of MicroRNAs in Regulation of Vascular Smooth Muscle Cell Differentiation**

Recent studies have demonstrated that microRNAs (miRNAs) are expressed in the vascular system and are involved in control of proliferation and differentiation of vascular smooth muscle cells. Each miRNA is able to regulate the expression of multiple target genes, frequently involved in the same cellular pathway. Conversely, target genes may be affected by more than 1 miRNA. It is presumed that thousands of human genes are targeted by miRNAs.36

Mature miRNAs are short, noncoding ribonucleic acid molecules, typically 22 nucleotides long, which bind to complementary sequences in the 3’ untranslated regions of target mRNA transcripts. PremiRNAs are exported from the nucleus to the cytoplasm, where they are processed by an RNase II enzyme, Dicer. They are then bound to the miRNA-induced silencing complex, which contains 2 key proteins, argonaute 2 and transactivation-responsive RNA-binding protein, to form mature miRNA.37 The mature miRNA and miRNA-induced silencing complex binds to complementary sites in the mRNA transcripts and negatively regulate gene expression. MiRNA binding leads to either mRNA degradation or inhibition of translation, depending on whether the miRNAs bind to their mRNA targets with perfect complementarity or imperfect complementarity, respectively.38,39 An important characteristic of miRNA expression is tissue- and cell-specific expression patterns.40

Overexpression of miR-145 or miR-143 is sufficient to promote differentiation and inhibit proliferation of cultured vascular smooth muscle cells.41,42 Deficiency of miR-145 or miR-143 promoted the synthetic phenotype of vascular smooth muscle, with increased migration to PDGF.43,44 miR-221 and miR-222, expression of which can be transcriptionally induced by PDGF, are also implicated in vascular smooth muscle cell differentiation.45,46 Overexpression of miR-221 reduced differentiation and increased proliferation and migration of vascular smooth muscle. Reduction of miR-221 increased expression of smooth muscle differentiation markers and blocked the effects of PDGF on proliferation and migration. miR-221 targets the 3’ untranslated regions of c-Kit and p27\(^{Kip1}\) mRNA.

In response to vascular injury, the expression of miRNA is dynamically regulated. At 7 days after balloon injury of the rat carotid artery, 113 of 140 artery miRNAs were differentially expressed; 60 were upregulated and 53 were downregulated.47 At 14 days after injury, 110 of 140 miRNAs were differentially expressed and at 28 days after injury, 102 of the 140 artery miRNAs were differentially expressed. In particular, miR-21 had more than 5-fold increase compared with control. Depletion of miR-21 caused decreased cell proliferation and increased cell apoptosis. Local delivery of an antisense oligonucleotide to knockdown miR-21 inhibited neointima formation in rat carotid artery after angioplasty.48 Adenoviral-mediated gene transfer of miR-145, which is downregulated after injury,47 inhibited neointimal lesion formation in injured rat carotid arteries.49 Surprisingly, neointimal formation was significantly impaired in miR-143 knockout, mir-145 knockout, or double knockout mice, which is contrary to other reports.48 Knockdown of miR-221 and miR-222, which are both increased in proliferative vascular smooth muscle, inhibits vascular smooth muscle cell proliferation and neointimal formation in rat carotid artery after injury.45

In endothelial cells, miR-126 has been shown to be required for migration and proliferation of human umbilical vein endothelial cells in response to vascular endothelial growth factor stimulation.49 miR-126 functions in part by directly repressing negative regulators of the vascular endothelial growth factor pathway, including the Sprouty-related protein SPRED1 and phosphoinositol-3 kinase regulatory subunit 2 (PIK3R2/p85-beta), and by regulating vascular cell adhesion molecule 1 expression, which mediates leukocyte adherence to endothelial cells.50

**Current Therapies Targeting Vascular Smooth Muscle Proliferation and Migration**

Currently available drug-eluting stent formulations deliver high concentrations of rapamycin, rapamycin analogues, or paclitaxel into the vessel wall (Figure 2). We will briefly review the mechanism of action of each.

**Rapamycin**

Rapamycin (AY-22,989, USAN name Sirolimus; Rapamune) is a macrocyclic triene antibiotic produced by the filamentous
activated by insulin.59

of growth factor and metabolic signaling and is strongly
inhibitors of mTORC1 and S6K1 and cell-type specific
Akt stands at the crossroads
RAPT (rapamycin target).53–56

studies of several species and has been termed mTOR, FRAP
(TOR) proteins to potently inhibit signaling to downstream
targets (Figure 2). A mammalian homologue has been cloned
from a soil sample from Easter Island (Rapa Nui).51

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mRNAs possessing a 5’ terminal oligopyrimidine tract
(5’TOP), a stretch of 4 to 14 pyrimidines found at the extreme
5’ terminus of some protein translation machinery mRNAs
(reviewed in Reference 60).60 Rapamycin has also been
shown to inhibit phosphorylation of 4E-BPs (eukaryotic
initiation factor 4E binding protein), which then can compete
with eIF4G proteins for binding with eIF4E.61 In addition,
rapamycin can also modulate the phosphorylation of eIF4B
and the translation elongation factor eEF2. Additional targets
involved in translational control may also be downstream of
TOR. Depletion of amino acids or glucose leads to rapid
dephosphorylation of 4E-BP1 (PHAS-I) and p70 S6 kinase
(S6K)-1.61

Rapamycin also significantly reduces the kinase activity of
the cyclin-dependent kinase (cdk) 4/cyclin D and cdk2/
cyclinE complexes. This is achieved by increasing the
cdk inhibitor p27Kip1 in various cell lines.62–64 The cdk inhibitor
p27Kip1 inhibits cyclin E/cdk2 kinase activity by forming a
complex (cyclinE/cdk2-p27Kip1), leading to inhibition of
phosphorylation of the retinoblastoma protein (pRb), as well
as inhibition of the dissociation of the pRb:E2F complex.
Loss of the ability to upregulate p27Kip1 in BC3H1 myogenic
cells is associated with rapamycin resistance and apoptosis on
serum withdrawal.65

Rapamycin inhibits intimal hyperplasia after both alloim-
mune and mechanical injury in rats.65,66 The mechanism of
this inhibition was demonstrated to involve a direct effect of
rapamycin on BC3H1 (myogenic cells)67 and smooth muscle
cell proliferation and migration.68,69 The inhibition of cellular
proliferation is associated with a reduction in the activity of
CDKs and in retinoblastoma protein phosphorylation (pRb),
leading to G1- to S-phase transition arrest.68 FK506 (tacrolimi-
us) and FK520 (an analog of FK506) failed to inhibit either
vascular smooth muscle cell proliferation or migration and in
fact antagonize rapamycin’s antiproliferative and antimigra-
tory properties through competition with FKBP12.68,69

Mitogen-induced downregulation of p27Kip1 is blocked by
rapamycin. In p27Kip1 null mice, we demonstrated relative
rapamycin resistance in mixed embryonic fibroblasts (MEF)
and splenic T-lymphocytes.63 In rapamycin-resistant myo-
genic cells (RR), constitutively low levels of p27Kip1 were
observed, which did not increase with serum withdrawal and
rapamycin. The lack of p27Kip1 reduces rapamycin-mediated
inhibition of smooth muscle cell migration, which suggests an
important role for p27Kip1 in the signaling pathway(s) regu-
lating smooth muscle cell migration.70,71 Silencing of p27Kip1
also blocked the inhibitory effects of rapamycin on migration
of endothelial cells.72 Decreased levels of p27Kip1 in the
vessel wall have been associated with markedly increased
neointimal response after wire injury.26 This study by the
Nabel group demonstrated that the increased neointimal
formation was due to lack of p27Kip1 in cells within the vessel
wall as well as bone marrow–derived cells. Thus, regulation
of p27Kip1 in nonvascular cells by rapamycin may be impor-
tant in modulating the arterial response to injury. There is
some controversy regarding the requirement for p27Kip1 in
mediating rapamycin’s inhibition of neointimal formation in
vivo, with 1 study reporting that neointimal formation in
p27Kip1 null mice was not enhanced after wire injury and that
rapamycin effectively reduced neointimal formation in
p27Kip1 null mice.73

Systemic (intramuscular) administration of rapamycin (0.5
mg/kg×3 days before injury and 0.25 mg/kg for 14 days) was
shown to significantly inhibit intimal proliferation in a porcine model of coronary artery balloon injury. Coronary arteries were analyzed at 4 weeks after injury revealing that rapamycin inhibited coronary intimal proliferation (control, 63±3.4% versus rapamycin, 36±4.5%, P<0.001). Rapamycin-coated stents were shown to inhibit restenosis in a porcine model of balloon injury/stent implantation. These preclinical data led to studies in humans demonstrating marked reduction of restenosis after rapamycin-coated stent implantation. Stents eluting rapamycin, including zotarolimus and everolimus, have been shown to be efficacious in reducing restenosis after stent implantation.

Diabetic patients may remain at increased risk of developing in-stent restenosis, despite the use of rapamycin-coated stents. Hyperleptinemia, which is seen in diabetes and metabolic syndrome, is a risk factor for stroke and progression of coronary artery disease. Leptin can promote platelet aggregation, inflammation, endothelial dysfunction, and vascular smooth muscle proliferation and migration, which are involved in atherogenesis. Leptin activates the mTOR-signaling pathway in primary murine vascular smooth muscle cells, stimulating proliferation in vitro. Exogenous leptin, administered at levels comparable to those found in obese humans, promoted neointimal hyperplasia and significantly increased the dose of rapamycin required for effective inhibition of neointimal formation (Figure 3). Combination therapy with a PI3K inhibitor, LY294002, and rapamycin effectively inhibited the leptin-enhanced neointimal hyperplasia. These data suggest that in the setting of hyperleptinemia, higher doses of an mTOR inhibitor or combination therapy with mTOR and PI3K inhibitors may be more efficacious in limiting neointimal formation.

As discussed above, reendothelialization is an important component of the healing process after vascular injury. Rapamycin has been shown to prevent vascular endothelial growth factor–mediated and serum-stimulated proliferation of endothelial cells in vitro. Additionally, rapamycin inhibits endothelial cell migration in a p27Kip1-dependent manner. The first-generation drug-eluting stents are highly effective at blocking neointimal hyperplasia but also exhibit reduced stent endothelialization. These observations may be due to rapamycin’s direct effects on endothelial proliferation and migration.

Paclitaxel
Paclitaxel was isolated from the bark of the Western yew tree in 1971. Paclitaxel induces cell cycle arrest in vascular smooth muscle and inhibits neointimal formation in animal models. Paclitaxel is extremely lipophilic, which promotes the rapid cellular uptake of the drug. Paclitaxel triggers the assembly of microtubules into extraordinarily stable yet disorganized and dysfunctional polymers. Unlike colchicine, which inhibits microtubule assembly, paclitaxel shifts the microtubule equilibrium toward assembly and polymerization. Several important biological processes are inhibited by paclitaxel, including mitotic spindle formation during cell division, intracellular transport, maintenance of cellular shape, and motility. Paclitaxel inhibits smooth muscle cell proliferation and migration in vitro and in vivo. Stents eluting paclitaxel reduced neointimal formation in a porcine coronary artery injury model. The arteries demonstrated incomplete healing with the late persistence of macrophages and fibrin deposition. These findings indicated the need for tightly controlled drug release and a narrow toxic-therapeutic window. Clinical trials with the paclitaxel-eluting stent have shown efficacy in preventing in-stent restenosis.

Concluding Remarks
Smooth muscle proliferation and migration after percutaneous intervention represent the end result of natural healing processes triggered by vascular injury. Vascular smooth muscle cell proliferation, especially after stent implantation, plays a critical role in neointimal hyperplasia through cellular expansion and extracellular matrix deposition. Elucidating the molecular mechanisms responsible for smooth muscle cell proliferation has led to the development of novel therapeutic approaches, including rapamycin- and paclitaxel-eluting stents that have significantly improved the care of patients with coronary artery disease. To address the concerns about the potentially increased incidence of stent thrombosis in patients treated with drug-eluting stents, newer stents and coronary devices have been developed such as drug-eluting stents with biodegradable polymers, drug-eluting stents that are polymer-free, stents with novel coatings, completely biodegradable stents, bifurcation stents, and drug-eluting balloons. Many of these are currently undergoing preclinical and clinical trials. Newer, potentially more efficacious, more specific, and safer approaches may be on the horizon.
Although targeting miRNAs represents a potential therapeutic strategy to specifically inhibit vascular smooth muscle proliferation and migration, to inhibit vascular inflammation, or to enhance vascular reendothelialization, additional work is required to identify precise targets and improve delivery.

Disclosures

None.

References


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